

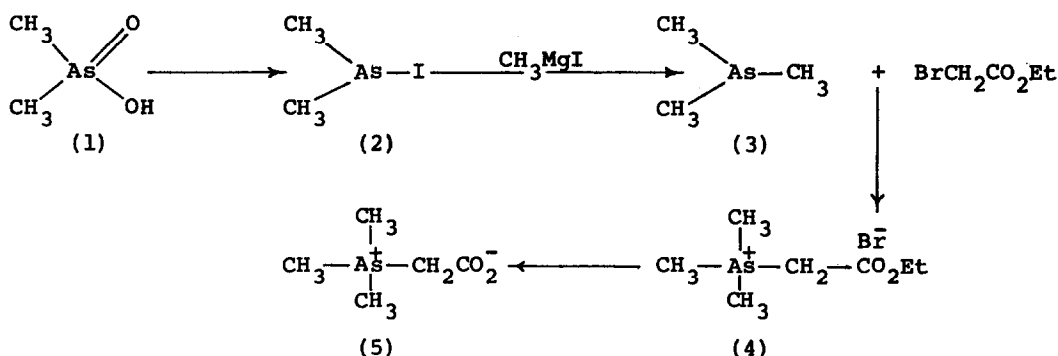
ISOLATION, CRYSTAL STRUCTURE AND SYNTHESIS OF ARSENOBETAINE, THE ARSENICAL CONSTITUENT OF THE WESTERN ROCK LOBSTER *PANULIRUS LONGIPES CYGNUS* GEORGE

John S. Edmonds and Kevin A. Francesconi
Western Australian Marine Research Laboratories, P.O. Box 20,
North Beach, Western Australia, 6020

Jack R. Cannon*, Colin L. Raston, Brian W. Skelton and Allan H. White
School of Chemistry, The University of Western Australia, Nedlands,
Western Australia, 6009

(Received in UK 8 March 1977; accepted for publication 21 March 1977)

It has been recognized for many years that appreciable quantities of arsenic (up to ~100 p.p.m.) occur in some edible marine animals¹. At present it seems that the arsenic compound(s) is acquired naturally and its presence does not necessarily reflect environmental pollution. Furthermore, the arsenic compound in sea food does not appear to be a toxic hazard for it is excreted rapidly by the human kidney². Although some progress has been made^{3,4} the precise structure of any organic arsenic compound present in a marine organism had not been elucidated when we began work. We now wish to report the isolation of arsenobetaine (5) from the tail muscle of the commercially-important western rock lobster (*Panulirus longipes cygnus* George). The nature of this substance has been elucidated by single crystal X-ray structure determination and its identity has been confirmed by comparison with a synthetic specimen obtained as follows.

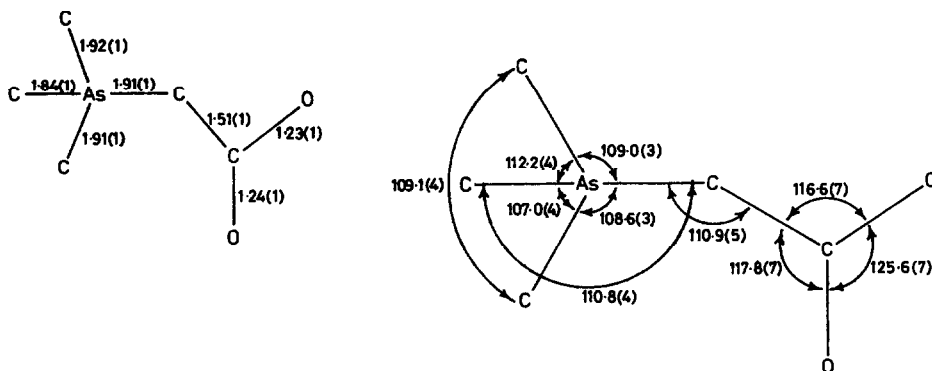


At all stages in the isolation procedure arsenic was located by vapour generation atomic absorption spectrometry⁵ following digestion of the sample with a mixture of perchloric and nitric acids. Homogenized tail muscle of the rock lobster (containing 26 p.p.m. of arsenic) was extracted with methanol, the methanol evaporated and the residue dissolved in water. Some

impurities were removed by extraction with ether. The remaining aqueous solution was acidified, shaken with phenol and the arsenic compound recovered from the phenol by dilution with ether followed by extraction with water. Further fractionation was achieved by passage of the aqueous solution through columns of Dowex 2X8(OH⁻) and Amberlite IRC50(H⁺). The eluate, which contained the arsenic compound, was then applied to a column of Zeokarb 225(H⁺) (SRC8, 2% DVB, 3.5 WR, <200 mesh); repeated chromatography involving careful elution of the column with dilute ammonia eventually gave a fraction containing 12.1% arsenic. This product gave two spots R_f 0.58 and 0.45 (revealed by exposure to iodine vapour) on thin layers of cellulose developed with *n*-butanol:acetic acid:water (60:15:25). Only the spot with R_f 0.58 contained arsenic and this product was isolated by preparative layer chromatography. After further chromatography on Zeokarb 225(H⁺) this fraction crystallized from acetone containing a trace of methanol.

Despite the fact that the solid product travelled as a single spot on a thin layer chromatogram, close inspection revealed that two crystal phases were present. Each of these was deliquescent, and it was found necessary to encapsulate the crystals in Canada balsam before mounting them on a diffractometer.

The first crystals were triclinic, $P\bar{1}$, $a = 8.780(4)$, $b = 7.088(3)$, $c = 6.914(3)\text{\AA}$; $\alpha = 80.70(3)$, $\beta = 77.54(3)$, $\gamma = 78.96(3)^\circ$; $D_m = 1.59(1)$, D_c ($Z = 2$) = 1.59g cm^{-3} . The structure was solved by the heavy atom method using 799 diffractometer reflections with $I > 3\sigma(I)$ and refined by full matrix least squares to a residual of 0.043. The thermal motions of all non-hydrogen atoms were refined anisotropically; all hydrogen atoms were located in a difference map and their positional parameters were successfully refined. The structure determination establishes the identity of the substance as $(\text{CH}_3)_3\text{As}^+\text{CH}_2\text{CO}_2^-\cdot\text{H}_2\text{O}$, the non-hydrogen molecular geometry being as follows (\AA , deg):



The hydrogen atoms of the water molecule bridge the oxygen atoms of successive carboxyl groups in an infinite one-dimensional array $O(H_2O) \dots O$ 2.75(1), 2.77(1)Å).

The second crystalline substance isolated from this mixture was also triclinic, $P\bar{1}$ or $P\bar{1}$, $a = 10.831(4)$, $b = 7.775(2)$, $c = 4.884(3)$ Å; $\alpha = 104.28(4)$, $\beta = 89.66(4)$, $\gamma = 110.37(3)^\circ$ but the diffractometer data reveal that it does not contain an arsenic atom or any other heavy atom; structural studies on this substance are proceeding.

Although arsenobetaine (5) has been the subject of a few biological studies⁶ we have been unable to find details of its synthesis. In the present work cacodylic acid (1) was converted into dimethyliodoarsine (2) after the method of Burrows and Turner⁷. Treatment of (2) with methylmagnesium iodide, as recommended by Challenger and Ellis⁸, gave trimethylarsine (3) which condensed smoothly with ethyl bromoacetate in benzene to afford the quaternary ester bromide (4). This substance crystallized from a large volume of acetone as prisms m.p. 143-145^o. Passage of an aqueous solution of (4) through a column of Dowex 2X8(OH⁻) followed by evaporation of the eluate gave arsenobetaine (5) which crystallized from acetone containing a little methanol as prisms of the monohydrate m.p. 204-210^o(dec.) (Found: C, 30.71; H, 6.64. $C_5H_{11}AsO_2 \cdot H_2O$ requires C, 30.63; H, 6.68%). The n.m.r. spectrum (D₂O) showed three singlets with δ 1.87 (9H, (CH₃)₃As), δ 3.28 (2H, CH₂) and δ 4.60 (HOD).

The natural and synthetic samples had the same R_f value and the crystallographic cell dimensions of the synthetic product agreed with those of the natural product within experimental error. The m.p. (203-210^o dec.) of selected crystals of the natural product was not depressed on admixture with synthetic arsenobetaine (5). The mass spectra (200^o/70eV) of the two samples were identical. No peaks with $m/e > 134$ appeared in the spectrum and the base peak was at m/e 103. In addition, both samples yielded a mixture of trimethylarsine and dimethylarsine in the ratio of ca. 30:70 after treatment first with strong aqueous sodium hydroxide and then with sodium borohydride³. The mechanism of this degradation has not yet been elucidated but it is possible that trimethylarsine oxide and cacodylic acid (1) are intermediates in this process.

Betaine esters are conveniently hydrolysed⁵ by treatment with a strongly basic anion exchange resin and as Dowex 2(OH⁻) was used during the isolation of arsenobetaine (5) from the extract of the rock lobster muscle the possibility that arsenobetaine (5) may be an artefact, derived by hydrolysis of a quaternary betaine ester or amide, was considered. However, we have

been unable to find any evidence to support this conjecture. Thin layer chromatography of a freshly-prepared methanol extract of the muscle, which had not been subjected to treatment with base, revealed that the arsenic-containing compound had the same R_f value (0.58) as synthetic arsenobetaine (5). Furthermore, chromatography of the quaternary betaine ester (4) under the same conditions showed it to be significantly more mobile (R_f 0.75) in this system.

Much work remains to be done on the biosynthesis and metabolism of arsenobetaine (5) in *Panulirus longipes cygnus* George, and possibly other marine organisms. However, the way is now clear for these studies to be undertaken and for a detailed toxicological assessment of this naturally occurring organic arsenic compound to be made.

We thank Dr. D.A. Hancock, Mr. P. Kerr, Mr. N.K. Roberts and Dr. S.B. Wild for helpful discussions.

References

1. A.C. Chapman, *Analyst*, 51, 548 (1926).
2. H.H. Schrenk and L. Schreibeis Jr., *Am. Ind. Hyg. Ass. J.*, 19, 225 (1958).
3. J.S. Edmonds and K.A. Francesconi, *Nature*, 265, 436 (1977).
4. W.R. Penrose, *CRC Critical Reviews in Environmental Control*, 4, 465 (1974).
5. L. Duncan and C.R. Parker, *Varian Techtron, Technical Topics* (Melbourne 1974).
6. A.D. Welch and R.L. Landau, *J. Biol. Chem.*, 144, 581 (1942) and references therein.
7. G.J. Burrows and E.E. Turner, *J. Chem. Soc.*, 117, 1373 (1920).
8. F. Challenger and L. Ellis, *J. Chem. Soc.*, 396, (1935).
9. E.W. Kosower and J.W. Patton, *J. Org. Chem.*, 26, 1318 (1961).